



Certificate of Analysis - Amended

Product Description	WA01 Oct4-eGFP Knock In	
Cell Line Provider	University of Wisconsin- Laboratory of Dr. James Thomson	
Lot Number	WA01(Oct4KI)-MCB-1	
Date Viald	04-October-2008	
Passage Number	p60	
Culture Platform	Feeder Independent	
	Media ¹ : TeSR	Matrix: Matrigel

The following testing specifications have been met for the specified product lot:


Test Description	Test Provider	Test Method	Test Specification	Result
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	≥ 15 Undifferentiated Colonies, ≤ 30% Differentiation	Pass
Identity by STR	UW Molecular Diagnostics Laboratory	PowerPlex 1.2 System by Promega	Consistent with known profile	Pass
Sterility - Direct transfer method	Apptec	30744	Negative	Pass
Mycoplasma	Bionique	M250	No contamination detected	Pass
Karyotype by G-banding	WiCell	SOP-CH-003	Normal karyotype	Pass

¹ These cells were cultured in the presence of G418 (Geneticin, Invitrogen catalog 11811) at a concentration of 50µg/ml. Not used for thawing, passaging, or freezing.

Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells.

Amendment(s):

Reason for Amendment	Date
CoA updated to include copyright information.	See signature
CoA updated for clarification of test specifications, added media footnote, and removed text regarding technical services	05-October-2010
CoA updated for format changes, clarification of test specifications, test method, addition of test provider, culture platform, and electronic signature, and reference to WiCell instead of the NSCB	19-August-2010
Original CoA	22-April-2009

Date of Lot Release	Quality Assurance Approval
22-April-2009	<div style="text-align: right;">12/30/2013</div> <div style="text-align: center;">  AMC Quality Assurance Signed by: XXXXXXXXXX </div>

Short Tandem Repeat Analysis*

Sample Report: **6618-STR**
WA01(Oct4KI)-MCB-1

UW HLA#: 59824

Sample Date: 11/06/08
Received Date: 11/07/08

Requestor: WiCell Research Institute

Test Date: 11/07/08

File Name: 081107

Report Date: 11/16/08

Sample Name: (label on tube) **6618-STR****Description:** DNA Extracted by WiCell
251.81 ng/ μ L; 260/280 = 1.91

Locus	Repeat #	STR Genotype
D16S539	5, 8-15	9,13
D7S820	6-14	8,12
D13S317	7-15	8,11
D5S818	7-15	9,11
CSF1PO	6-15	12,13
TPOX	6-13	8,11
Amelogenin	NA	X,Y
TH01	5-11	9.3,9.3
vWA	11, 13-21	15,17

Comments: Based on the 6618-STR DNA submitted by WI Cell dated 11/06/08 and received on 11/07/08, this sample (UW HLA# 59824) matches exactly the STR profile of the human stem cell line **H1** comprising 15 allelic polymorphisms across the 8 STR loci analyzed. No STR polymorphisms other than those corresponding to the human H1 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. This result suggest that the 6618-STR DNA sample submitted corresponds to the H1 stem cell line and was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells. Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is ~5%. These results were communicated via phone to the Cytogenetics laboratory of the WiCell Research Institute on Monday, October 27, 2008. A preliminary copy of this report was issued via electronic mail to the WI Cell Research Institute on Monday, November 17, 2008.

* Testing to assess engraftment following bone marrow transplantation was accomplished by analysis of human genetic polymorphisms at STR loci. This methodology has not yet been approved by the FDA and is for investigational use only.

Test Facility:

This report is confidential. No part may be used for advertising or public announcement without written permission. Results apply only to the sample(s) tested.

Report Number
795304
Page 4 of 7

WiCell Research Institute

December 16, 2008
P.O. #: _____

STERILITY TEST REPORT

Sample Information: hES Cells
3: WA01 (Oct4KI)-MCB-1

Date Received: November 25, 2008
Date in Test: December 01, 2008
Date Completed: December 15, 2008

Test Information: Test Codes: 30744, 30744A
Immersion, USP / 21 CFR 610.12
Procedure #: BS210WCR.201 (Modified: Alternate media used.)

TEST PARAMETERS	PRODUCT	
Approximate Volume Tested	0.5 mL	0.5 mL
Number Tested	2	2
Type of Media	SCD	FTM-T-L-S
Media Volume	400 mL	400 mL
Incubation Period	14 Days	14 Days
Incubation Temperature	20 °C to 25 °C	30 °C to 35 °C
RESULTS	2 NEGATIVE	2 NEGATIVE

QA Reviewed: _____
Page 1 Signed

Reviewed: _____
Page 1 Signed

Testing conducted in accordance with current Good Manufacturing Practices.



Document#: DCF3013D
 Edition#: 10
 Effective Date: 07/15/2003
 Title: **M-250 FINAL REPORT SHEET**

M-250 FINAL REPORT

Direct Specimen Culture
 Procedure 3008, 3011, 3013

TO: **Wicell QA**

BTL SAMPLE ID#: **55226** P.O.#: DATE REC'D: **11/04/2008**

TEST/CONTROL ARTICLE:

6618 WA01 (OCT4 KI) MCB1

LOT#: **NA**

DIRECT CULTURE SET-UP (DAY 0)

DATE: **11/05/2008**

INDICATOR CELL LINE (VERO)

SEE DNA FLUOROCHROME RECORD SHEET

				DATE
THIOGLYCOLLATE BROTH	DAY 7	+	⊖	<u>11/12/2008</u>
	DAY 28	+	⊖	<u>12/03/2008</u>
BROTH-FORTIFIED COMMERCIAL				
<u>0.5</u> mL SAMPLE	DAY 7	+	⊖	<u>11/12/2008</u>
<u>6.0</u> mL BROTH	DAY 28	+	⊖	<u>12/03/2008</u>
BROTH-MODIFIED HAYFLICK				
<u>0.5</u> mL SAMPLE	DAY 7	+	⊖	<u>11/12/2008</u>
<u>6.0</u> mL BROTH	DAY 28	+	⊖	<u>12/03/2008</u>
BROTH-HEART INFUSION				
<u>0.5</u> mL SAMPLE	DAY 7	+	⊖	<u>11/12/2008</u>
<u>6.0</u> mL BROTH	DAY 28	+	⊖	<u>12/03/2008</u>

(See Reverse)

Document#: DCF3013D
 Edition#: 10
 Effective Date: 07/15/2003
 Title: M-250 FINAL REPORT SHEET

SAMPLE ID#:	55226	AEROBIC	MICROAEROPHILIC	DATE
AGAR PLATES-FORTIFIED COMMERCIAL	DAY 7	+ ⊖	+ ⊖	<u>11/12/2008</u>
	DAY 14	+ ⊖	+ ⊖	<u>11/19/2008</u>
	DAY 21	+ ⊖	+ ⊖	<u>11/26/2008</u>
AGAR PLATES-MODIFIED HAYFLICK	DAY 7	+ ⊖	+ ⊖	<u>11/12/2008</u>
	DAY 14	+ ⊖	+ ⊖	<u>11/19/2008</u>
	DAY 21	+ ⊖	+ ⊖	<u>11/26/2008</u>
AGAR PLATES-HEART INFUSION	DAY 7	+ ⊖	+ ⊖	<u>11/12/2008</u>
	DAY 14	+ ⊖	+ ⊖	<u>11/19/2008</u>
	DAY 21	+ ⊖	+ ⊖	<u>11/26/2008</u>
BROTH SUBCULTURES (DAY 7)		DATE: <u>11/12/2008</u>		
AGAR PLATES-FORTIFIED COMMERCIAL	DAY 7	+ ⊖	+ ⊖	<u>11/19/2008</u>
	DAY 14	+ ⊖	+ ⊖	<u>11/26/2008</u>
	DAY 21	+ ⊖	+ ⊖	<u>12/03/2008</u>
AGAR PLATES-MODIFIED HAYFLICK	DAY 7	+ ⊖	+ ⊖	<u>11/19/2008</u>
	DAY 14	+ ⊖	+ ⊖	<u>11/26/2008</u>
	DAY 21	+ ⊖	+ ⊖	<u>12/03/2008</u>
AGAR PLATES-HEART INFUSION	DAY 7	+ ⊖	+ ⊖	<u>11/19/2008</u>
	DAY 14	+ ⊖	+ ⊖	<u>11/26/2008</u>
	DAY 21	+ ⊖	+ ⊖	<u>12/03/2008</u>

RESULTS: No detectable mycoplasmal contamination

12/3/08
Date

Laboratory Director

M-250 Procedural Summary: The objective of this test is to ascertain whether or not detectable mycoplasmas are present in an *in vitro* cell culture sample, be it a primary culture, hybridoma, master seed stock or cell line. This procedure combines an indirect DNA staining approach to detect non-cultivable mycoplasmas with a direct culture methodology utilizing three different mycoplasma media formulations. The indirect approach involves the inoculation of the sample into a mycoplasma-free VERO (ATCC) indicator cell line and performing a DNA fluorochrome assay after 72-120 hours of incubation. The direct culture aspect of the test utilizes three different mycoplasma media including both broth and agar formulations. The sample is inoculated into each of the 3 broth formulations and also onto duplicate plates (0.1 mL/plate) for each of the 3 agar formulations. Subculture from broth to fresh agar plates is carried out after 7 days incubation. Agar plates are incubated aerobically and microaerophilically in order to detect any colony forming units morphologically indicative of mycoplasma contamination. Issuance of the final report with signature of the Scientific Director/Study Director signifies that the required controls were performed concurrently with the test sample(s) as detailed in the referenced SOPs and that all test conditions have been found to meet the required acceptance criteria for a valid test, including the appropriate results for the positive and negative controls.



APPENDIX I

Document #: DCF3008A
Edition #: 06
Effective date: 9/17/2003
Title: DNA FLUOROCHROME ASSAY RESULTS

DNA-FLUROCHROME ASSAY RESULTS

Procedures 3008, 3009, 3011

Sample ID # 55226 M-250 Date Rec'd: 11/04/2008 P.O. #

Indicator Cells Inoculated: Date/Initials: 11/6/08 / JA

Fixation: Date/Initials: 11/10/08 / KG

Staining: Date/Initials: 11/10/08 / KG

TEST/CONTROL ARTICLE:

6618 WA01 (OCT4 KI) MCB1

LOT# NA

Wicell QA
WiCell Research Institute

DNA FLUROCHROME ASSAY RESULTS:

X **NEGATIVE:** A reaction with staining limited to the nuclear region, which indicates no mycoplasmal contamination.

 POSITIVE: A significant amount of extranuclear staining which strongly suggests mycoplasmal contamination.

 INCONCLUSIVE:

 A significant amount of extranuclear staining consistent with low - level mycoplasmal contamination or nuclear degeneration.

 A significant amount of extranuclear staining consistent with bacterial, fungal or other microbial contaminant or viral CPE. Morphology not consistent for mycoplasmal contamination.

COMMENTS: _____

Date: 11/10/08 Results Read by: KG Date of Review: 11/10/08 Reviewed by: SA

Report Date: November 11, 2008

Case Details:

Cell Line: WA01(Oct4KI)-MCB-1 (6618)

Passage #: 62

Date Completed: 11/10/2008

Cell Line Gender: Male

Investigator: WiCell Stem Cell Bank

Specimen: hESC on Matrigel

Date of Sample: 10/31/2008

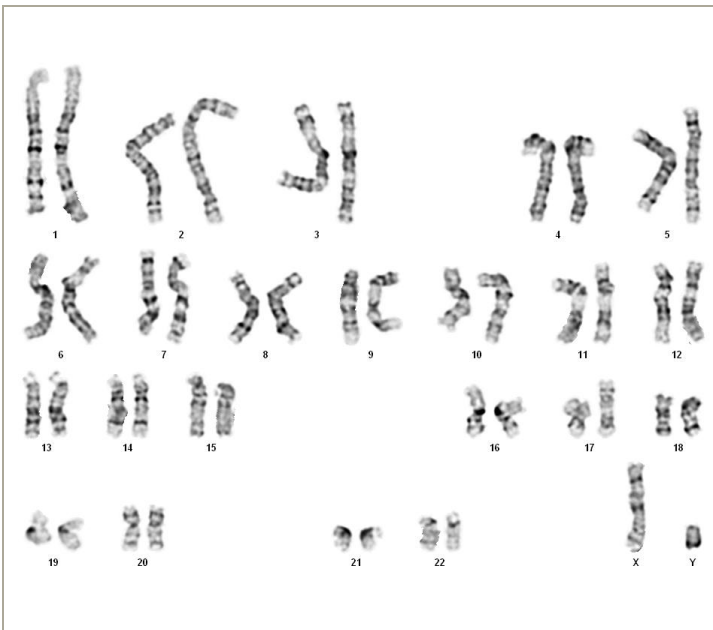
Tests, Reason for: Wisc Bank- FTDL

Results: 46,XY

Completed by ST, CLSp(CG), on 11/10/2008

Reviewed and interpreted by KDM, PhD, FACMG, on 11/10/2008

Interpretation: No abnormalities were detected at the stated band level of resolution.



Cell: S01-03

Slide: B

Slide Type: Karyotyping

Cell Results: Karyotype: 46,XY

of Cells Counted: 20

of Cells Karyotyped: 3

of Cells Analyzed: 7

Band Level: 450-550

Results Transmitted by Fax / Email / Post
Sent By: _____

Date: _____
Sent To: _____